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population of mould and yeast as well as the total aerobic bacteria of the combined-treated grains (60°C, 4.0 kGy) remained nearly the same (i.e. 5.0 and 4.3 log cycles reduction, respectively). The control of the moist heat-treated grains, however, had mould and yeast and total aerobic bacteria counts lowered by 1.5 and 1.3 log cycles, respectively, after three months' storage at 80% r.h. The grains did not become rancid.

Triplicate samples showed that only control grains (20L and 20H) and the grains (20H) irradiated with 4.0 kGy contained 0.8–4.0 µg/kg of aflatoxin B₁ after three months' storage at 80% r.h. and 28°C.

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DETERMINATION OF IRRADIATION D-VALUES FOR *Aeromonas hydrophila* IN GROWTH MEDIUM, BUFFER AND FISH

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To assess the potential of irradiation processing as a means of controlling the presence of *Aeromonas hydrophila* in marine products [1] three clinical isolates (K144, BA2 and BW83) and two food isolates (F6-10 and B2-10) were used in these studies. The cultures were irradiated in a caesium-137 source at doses up to 125 krad. Cultures were irradiated directly at 2 or 22°C in BHI growth medium, in potassium phosphate buffer (0.1M, pH 7.2), or in ground blue fish (Table I).¹ The number of survivors after exposure to various irradiation doses was determined by plating appropriate dilutions on duplicate plates of phenol red starch agar with 10 mg/L ampicillin, and enumerating amylase⁺ colonies after 24 h incubation at 28°C. Survivor plots (log₁₀ number of survivors versus dose) were determined by regression analysis of the data; correlation coefficients ≥0.96 were obtained for all strains and variables. Decimal reduction doses (D values in krad)² were calculated as the reciprocal of the slope obtained from the regression analysis.

The D-values observed with the different strains were determined (Tables II–IV). Comparison of our data with those of Tarkowski et al. [2]

¹ BHI = Brain, heart, infusion.

² 1 rad = 1.00 × 10⁻² Gy.

TABLE I. EFFECT OF GROWTH PHASE ON D-VALUES FOR *A. hydrophila* (irradiated in culture broth and plated on nutrient agar)

Strain	Stationary phase cells	log phase cells
K144	18.1	18.0
BA2	19.0	19.5
BW83	16.5	18.3
F6-10	17.6	16.9
B2-10	17.8	21.6

TABLE II. EFFECT OF PLATING MEDIUM AND IRRADIATION MEDIUM ON D-VALUES FOR *A. hydrophila*

Strain	Starch ampicillin agar		Nutrient agar	
	Growth medium	Phosphate buffer	Growth medium	Phosphate buffer
K144	16.2	15.5	15.8	14.8
BA2	18.7	18.1	18.8	18.6
BW83	16.8	15.9	15.7	15.6
F6-10	15.7	15.7	15.5	14.0
B2-10	15.5	13.7	15.4	14.9

TABLE III. EFFECT OF TEMPERATURE OF IRRADIATION ON D-VALUES OF *A. hydrophila* IN FISH (plated on starch ampicillin agar)

Strain	Temperature of irradiation (°C)		
	22	2	-15
K144	13.7	17.7	26.2
BA2	15.2	19.3	31.4
BW83	14.5	16.1	34.0
F6-10	11.0	14.1	23.3
B2-10	11.3	15.6	22.2

TABLE IV. D-VALUES OF *A. hydrophila*
IRRADIATED IN GROUND BEEF AT 2°C
(plated on starch ampicillin agar)

Strain	
K144	14.0
BA2	14.3
BW83	18.9
F6-10	15.1
B2-10	15.0

indicate that *A. hydrophila* is slightly more radiation resistant than *Yersinia enterocolitica* and *Campylobacter jejuni*, but not as resistant as *Salmonella* when these pathogens were irradiated in raw beef. However, our D-values for *A. hydrophila* in fish at 2°C are similar to those reported by Lambert and Maxey [3] for *C. jejuni* in ground beef and turkey. Overall, the results of our study indicate that a dose of 100 krad should be efficacious for the elimination of the levels of *A. hydrophila* encountered in retail fresh foods.

REFERENCES

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